

A MULTI-CENTRIC PROSPECTIVE STUDY ON CANDIDIASIS- THE CLINICO-EPIDEMIOLOGICAL ASPECT AND ANTIFUNGAL SUSCEPTIBILITY PATTERN IN TERTIARY CARE HOSPITALS

Atreyi Chakraborty¹, Sampurna Biswas Pramanik²

Received : 30/11/2024
Received in revised form : 09/01/2025
Accepted : 31/01/2025

Keywords:

Candida species, antifungal susceptibility, Amphotericine B resistance, Micafungin susceptibility, non albicans Candida, Immunosuppression.

Corresponding Author:

Dr. Atreyi Chakraborty,
Email: dratreyi10@gmail.com

DOI: 10.47009/jamp.2025.7.6.101

Source of Support: Nil,
Conflict of Interest: None declared

Int J Acad Med Pharm
2025; 7 (6); 536-542



¹Assistant Professor, Department of Microbiology, Medical College, Kolkata, West Bengal, India
²Associate Professor, Department of Microbiology, School of Tropical Medicine, Kolkata, West Bengal, India

ABSTRACT

Background: The purpose of the study was to determine the prevalence of different *Candida* species causing infections and their antifungal susceptibility pattern. **Materials and Methods:** Various types of clinical specimen collected from the patients with suspected fungal infection were examined by wet mount preparation and gram staining. Specimens were subsequently cultured on SDCA and incubated at 25°C and 37°C aerobically for 4 weeks. *Candida* isolates thus identified were subcultured on CHROM agar and corn meal agar for speciation. Confirmation of species level identification and antifungal susceptibility pattern were obtained using automated VITEK 2 method. **Result:** 221 culture positive *Candida* isolates were obtained from 3371 non-repetitive consecutive clinical specimens collected from suspected cases of cutaneous or invasive systemic fungal infections. 16.89%, 12.61%, 12.31%, 9.56%, 7.5% and 2.88% of the tongue scrapings, sputum, nail clippings, urine, esophageal biopsies and blood samples yielded growth of *Candida*. *Candida albicans* accounted for 57.92% isolates followed by *Candida tropicalis* (14.03%), *C. auris* (5.88%), *C. parapsilosis* (5.43%) and *C. glabrata* (5.43%). The study highlighted on different clinico-epidemiological aspects. 61.09% of Candidiasis patients were male. Candidiasis was most common among 18-45 years (36.80%). Immunosuppression due to HIV turned out to be the most significant underlying disease. Diabetes mellitus was more common in non-*albicans* Candidiasis. *Candida albicans* was completely susceptible in vitro to Voriconazole, Caspofungin and Micafungin. Amphotericine B (24.21%) and Fluconazole (7.81%) were most in-vitro resistant against *Candida albicans*. Fluconazole, Amphotericine B and Caspofungin were resistant in 36.56%, 29.03% and 20.43% non *albicans* isolates respectively. Voriconazole (9.68%) and Flucytosine (12.9%) have also reported an increasing trend of resistance against these isolates. **Conclusion:** Echinocandins namely Micafungin was the only drug showing 100% efficacy against *Candida* isolates. Amphotericine B and fluconazole, flucytosine exhibited increasing resistance against *Candida albicans*. Caspofungin resistance is alarming as not only a few Flucanazole resistant isolates are showing complete or intermediate resistance to Caspofungin in-vitro but increasing MIC values are observed in few Fluconazole sensitive isolates also. Resistance of non *albicans* *Candida* against both Azoles and Echinocandin are increasing probably due to an emerging trend of using either of them as first line therapy in invasive infections. Judicious use of these drugs in invasive and nosocomial candidiasis is therefore the need of the hour to prevent rapid emergence of resistance.

INTRODUCTION

Fungal infections are an important cause of mortality and morbidity worldwide, more so in immunocompromised patients. Candidiasis is the most common opportunistic fungal infection reported

worldwide being ubiquitous in their presence in the environment. *Candida* can cause a wide range of diseases from superficial localized cutaneous infections to severe disseminated invasive clinical infections. *Candida* is considered as part of normal microbial flora of oral cavity, gastro-intestinal and

genitor-urinary tract in a considerable portion of healthy adult population and it may result in colonization and opportunistic infection in persons with significant predisposing factors and co-morbidities, most significant being immune-compromised individuals. *Candida albicans* still remains the most common *Candida* species causing human infections but quite a large proportion of infections are now reportedly to be due to non-*albicans* species of *Candida*. There is an emerging trend of in-vitro resistance towards antifungal drugs commonly used among both *albicans* and non-*albicans* species of *Candida*. This changing trend of *Candida* infections calls for further investigation. Therefore our study was undertaken to determine prevalence of different *Candida* species causing infections and antifungal susceptibility pattern of those isolates.

MATERIALS AND METHODS

After obtaining approval from Institutional Ethics Committee, the multicentric prospective observational study was conducted at the Department of Microbiology of two tertiary care hospitals in Eastern India for six months from April 2024 to September 2024. Consecutive, non-repetitive samples were collected from the patients presenting to us with suspected fungal infection during the study period. Different types of clinical specimens namely sputum, tracheal aspirate, broncho-alveolar lavage, tissue biopsy specimens, nail clippings, cutaneous scrapings, tongue scrapings, blood and body fluids such as ascitic fluid, pleural fluid, and urine were collected from the patients after obtaining informed consent. Detailed history regarding the present illness and co-morbidities was recorded. Data regarding the HIV seropositivity status of the patient was recorded with informed consent maintaining utmost confidentiality.

The specimens were primarily examined by direct examination of wet mount preparation and gram-stained smear. Sputum specimens with Barlett score >1 was accepted for further processing by sub-culture on Sabouraud dextrose chloramphenicol agar (SDCA) media. Presence of *Candida* isolates as pathogen in urine were confirmed by repeated isolation of the same in urine of patients with a colony count of 10⁵ CFU/ml or higher on at least one occasion with symptomatic UTI. Direct microscopic findings were observed in the wet mount preparations after treating the specimen with 10% KOH and findings in the form of pseudohyphae or budding yeast were recorded. Then the specimens were cultured on SDCA (Sabouraud dextrose chloramphenicol agar) media and incubated at 25°C and 37°C aerobically for 4 weeks and observed for the growth on every alternate day during the first week and twice a week for next 3 weeks. Gram-stained smears from the colonies were prepared once growth of pasty and cream coloured colonies observed. Presence of Gram positive single or

budding yeast cell with or without pseudohyphae was recorded. Germ tube test at 37°C was performed to differentiate *Candida albicans* and *Candida dublinensis* from other *Candida* species. Specimen identified as *Candida* isolates were subsequently subcultured on CHROMagar media and corn meal agar for speciation. Confirmation of species level identification along with antifungal susceptibility pattern was obtained using automated VITEK2 method.

RESULTS

221 culture positive *Candida* isolates were obtained from a total number of 3371 non-repetitive consecutive clinical specimens collected for fungal culture in our Mycology laboratory. An overall culture positivity rate of 6.56% of *Candida* infection was observed in this tertiary care hospital inclusive of both cutaneous and invasive systemic fungal infections. Highest percentage (16.89%) of *Candida* isolates was reported as a causative pathogen from tongue scraping specimen after due consideration of the associated clinical history in each patient. 12.61% of the sputum specimen also yielded growth of *Candida* species. Cases of onychomycoses caused by *Candida* species accounted for 12.31% of the nail clipping specimen. *Candida* isolates were associated with 9.56% & 7.5% of UTI and esophagitis cases respectively. *Candidaemia* was reported in 2.88% isolates from aerobically incubated blood culture bottle [Table 1].

Candida albicans was the most important species responsible for causing majority of the (57.92%) fungal infections caused by *Candida* in our study. *Candida tropicalis* was the most frequently isolated non-*albicans* species accounting for 14.03% of all *Candida* infections. Emergence of *Candida auris* and *Candida glabrata* as the next most frequent causes of non-*albicans* *Candida* infections raises concern because of their well-documented antifungal resistance [Table 2 & 3].

Infections due to *Candida* was most common among persons aged 18-45 years. 14.48% of the isolates were from patients aged below 18 years and non-*albicans* was responsible for most of the cases in the paediatric age group [Table 4]. Candidiasis was more common (61.09%) in male patients and two-third of the cases in them were caused by *C. albicans* only whereas non-*albicans* *Candida* species were responsible for 51.61% cases in female patients who accounted for 38.91% of total *Candida* infections [Table 5].

Immunosuppression due to HIV turned out to be the single most significant underlying disease condition favouring fungal infection due to *Candida* species irrespective of their CD4 T lymphocyte count or whether the patient is on ART therapy or not and the isolates mostly belonged to *C. albicans* in these patients. *Candida albicans* was the most frequent isolate in presence of other associated disease conditions known for lowering the immune

competence such as Tuberculosis, malignancy, auto-immune diseases and chronic kidney disease. However, Diabetes, the second most frequently associated co-morbidity in cases of Candidiasis was more common in patients infected with non-albicans Candida species [Table 6].

Candida albicans isolates have shown highest percentage of in-vitro resistance towards Amphotericine B (24.21%) followed by Fluconazole. 7.81% of *Candida albicans* isolates were resistant to Fluconazole and 9.34% isolates were susceptible only to higher concentration of the drug. All the *C. albicans* isolates were completely susceptible in vitro to Voriconazole, Caspofungin and Micafungin [Table 7]. Non albicans isolates of *Candida* have demonstrated a higher degree of in-vitro resistance in comparison with *C. albicans* to all the antifungals tested for except Micafungin. All the isolates were completely susceptible towards Micafungin only. Fluconazole was the most resistant (36.56%) antifungal drug against Non albicans *Candida* isolates closely followed by Amphotericine B and Caspofungin, resistant in vitro against 29.03% and 20.43% of NAC isolates respectively. Voriconazole (9.68%) and Flucytosine (12.9%) have also reported an increasing trend respectively of resistance against NAC isolates [Table 8].

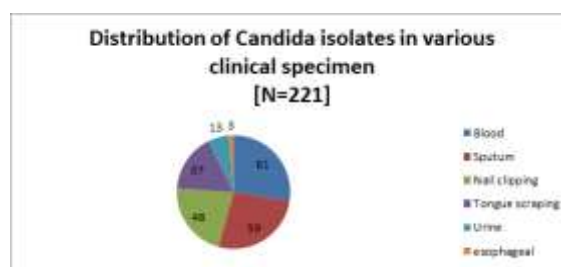


Figure 1: Distribution of candida isolates in various clinical specimen

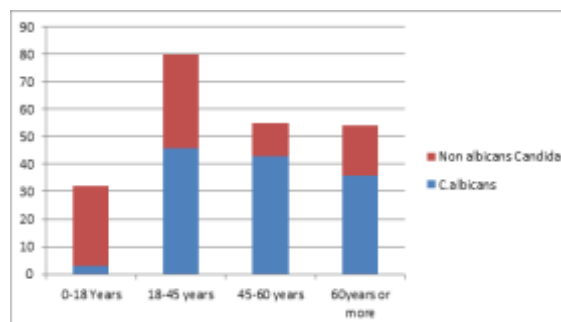


Figure 2: Distribution of candida albicans and non albicans species among culture positive samples in various age groups.

Table 1: Incidence of Candida isolates in various clinical specimen.

Specimen	Total number of specimen collected(N=3371)	Growth obtained (N=221)	Percentage(%) positivity among each type of specimen obtained
Blood	2118	61	2.88
Sputum	468	59	12.61
Nail clipping	390	48	12.31
Tongue scraping	219	37	16.89
Urine	136	13	9.56
Esophageal brushing/biopsy	40	3	7.5

Table 2: Distribution of Candida albicans and non albicans species causing infection in different clinical specimen in tertiary care hospital

Specimen(N=221)	Candida albicans(N=128)	Candida non-albicans(N=93)
Blood	24	37
Sputum	43	16
Nail clipping	19	29
Tongue scraping	37	0
Urine	2	11
Esophageal	3	0

Table 3: Age wise distribution of Candidiasis cases (Candida albicans-CA, Non albicans Candida species-NCA)

Age group	Total (N=221)	%
0-18	32	14.48
	CA	3
	NCA	29
18-45	80	36.20
	CA	46
	NCA	34
45-60	55	24.89
	CA	43
	NCA	12
60-rest	54	24.43
	CA	36
	NCA	18

Table 4: Gender wise distribution of Candidiasis cases

Gender	Total (N=221)		%
Male	135		61.09
	CA	90	
	NCA	45	
Female	86		38.91
	CA	38	
	NCA	48	

Table 5: Co-morbidities associated with cases of Candidiasis

Predisposing Factors	Total (N=221)		%
HIV reactive	52		23.53
	CA	42	
	NCA	10	
Diabetes Mellitus	51		23.07
	CA	19	
	NCA	32	
Malignancy	25		11.31
	CA	18	
	NCA	7	
Tuberculosis	19		8.59
	CA	14	
	NCA	5	
Auto-immune diseases	22		9.95
	CA	18	
	NCA	4	
Chronic renal disease	6		2.71
	CA	3	
	NCA	3	

Table 6: Species wise distribution of Candida isolates causing infection

Micro-organism	N=221	Percentage (%)
Candida albicans	128	57.92
Candida auris	13	5.88
Candida parapsilosis	12	5.43
Candida Ciferri	9	4.07
Candida glabrata	12	5.43
Candida tropicalis	31	14.03
Candida famata	5	2.26
Candida krusei	2	0.90
Candida gulermondii	5	2.26
Candida dublinensis	2	0.90
Candida lipolytica	1	0.45
Candida luzitaniae	1	0.45

Table 7: In-vitro susceptibility pattern of commonly used antifungals against Candida albicans isolates(n=128)

Name of the antibiotic	Sensitive		Intermediate Sensitive		Resistant	
	Number of isolates	Percentage	Number of isolates	Percentage	Number of isolates	Percentage
Fluconazole	106	82.81	12(SDD)	9.34	10	7.81
Voriconazole	128	100	0	0	0	0
Caspofungin	117	90.41	11	8.59	0	0
Micofungin	128	100	0	0	0	0
Amphotericin B	97	75.78	0	0	31	24.21
Flucytocin	125	97.66	0	0	3	2.34

Table 8: In-vitro susceptibility pattern of commonly used antifungals against NAC isolates(n=93)

Name of the antibiotic	Sensitive	Percentage	Resistant	Percentage
Fluconazole	59	63.44	34	36.56
Voriconazole	84	90.32	9	9.68
Caspofungin	74	79.57	19	20.43
Micofungin	93	100	0	0
Amphotericin B	66	70.97	27	29.03
Flucytocin	81	87.1	12	12.9

Table 9: In-vitro resistance pattern of commonly used antifungals against different NAC isolates(n=93)

Micro-organism	Fluconazole (N=34)	Voriconazole (N=9)	Caspofungin (N=19)	Amphotericin B (N=27)	Flucytocin (N=12)
Candida tropicalis(n=31)	6	2	0	4	0
Candida auris(n=13)	9	2	2	7	1

<i>Candida parapsilosis</i> (n=12)	2	0	0	5	1
<i>Candida glabrata</i> (n=12)	5	2	5	9	4
<i>Candida ciferri</i> (n=9)	9	3	9	0	4
<i>Candida famata</i> (n=5)	1	0	1	0	0
<i>Candida guillemondii</i> (n=5)	0	0	2	0	0
<i>Candida krusei</i> (n=2)	2	0	0	2	2
<i>Candida dublinensis</i> (n=2)	0	0	0	0	0
<i>Candida lipolytica</i> (n=1)	0	0	0	0	0
<i>Candida lusitanae</i> (n=1)	0	0	0	0	0

Note- All the NAC isolates were in-vitro susceptible against Micafungin, hence the drug was not included in this table.

N – denotes the total number of resistant NAC isolates against the particular drug

n- denotes the total number of isolates of the particular *Candida* species.

DISCUSSION

The spectrum of the disease caused by *Candida* varied from localised onychomycoses and oropharyngeal candidiasis, urinary tract infection to more severe and invasive systemic ones such as fungal pneumonia and candidemia. Invasive Candidiasis in form of persistent Candidemia was the commonest type of infection being closely followed by fungal pneumonia in our study. Non albicans species were responsible for majority of the cases of candidemia in this study corroborating with other studies reporting a shifting trend acknowledged in most of the continents globally.^[1] Being an important cause of health care associated infection characterised by high morbidity and mortality rate it raises a matter of concern. Non-albicans *Candida* isolates were more frequently obtained from patients suffering from onychomycoses and urinary tract infection as well corroborating with findings obtained from other studies. Taci et al study suggested an increasing trend of empirical antifungals usage such as azoles, use of indwelling medical devices, malignancies and long term immunosuppressive therapy to be associated with such infections.^[2] *Candida tropicalis*, *C.auris*, *C.glabrata* and *C.parapsilosis* emerged as the most frequently isolated NAC in our study. Recent studies by Ortiz et al,^[3] (2022) and Megri et al,^[4] (2020) also reported surge in infections by those candida species well known for their resistance towards azoles and echinocandins. NM Reda et al,^[5] study done in Egypt in 2023 reported NAC isolates in more than half of the candidemia patients in paediatric age group and *C.tropicalis* and *C.parapsilosis* being the most frequent ones. Predominance of NAC isolates was also observed in patients below 18 years in our study though *C.albicans* was responsible for majority of the cases in all other age groups. Incidences of candidiasis were most frequent among patient aged more than 45 years with a male preponderance correlating with the findings of other studies across the globe including Indian ones.^[6-8] An Indian study exclusively carried out in HIV seropositive individuals by Maheswari et al in 2016 detailed that *C.albicans* continued to be the commonest fungal pathogen in those patients. Findings of our study also corroborated that.^[9] NAC isolates prevalent among HIV seropositive cases mostly belonged to

C.tropicalis and *C.glabrata*. Sero-positivity for HIV irrespective of the CD4 count of the cases and Diabetes mellitus were most significant predisposing factors. Immunosuppression either due to malignancy or use of immune-modulators in autoimmune diseases was also frequently found to be the underlying cause for candidiasis. Co-morbidities such as active cases of tuberculosis and chronic kidney disease were also associated with fungal disease though less frequently. *Candida albicans* was the most common isolate in patients with significant co-morbidities except Diabetes mellitus. NCA isolates mostly belonging to *C.tropicalis* constituted major portion of candidiasis cases in diabetics. *C.glabrata*, *C.krusei* and *C.auris* were also isolated in quite a few cases in diabetics. Chouhan et al study carried out in Bhopal in 2019 indicated a higher carriage rate of NCA species such as *C.glabrata* & *C.tropicalis* in oral cavity of diabetics than *C.albicans*.^[10] The factors favouring local or systemic candidiasis in diabetics are attributed to several mechanisms,^[11] such as increased adhesion and colonization of pathogen to epithelial surface,^[12] increased salivary and blood glucose concentration,^[13] increased microvascular degeneration and ineffective microbiocidal activities of PMNs, macrophage and lymphocytes.^[11] Increased ability for biofilm production mainly by *C.glabrata* and also by *C.tropicalis* and *C.albicans* are probably responsible for their frequent association with infection in diabetics.^[14] In corroboration with other studies carried out in last decade, ours also indicated a changing pattern in the species prevalence of *Candida*. Though *C.albicans* continues to be responsible for majority of the cases reported, yet the incidences of NAC namely *C.tropicalis*, *C.auris*, *C.parapsilosis*, *C.glabrata* and *C.ciferri* has increased nowadays.^[15] A 2016 study from Kolkata reported *C.tropicalis*, *C.haemulonii* and *C.glabrata* as most prevalent ones among NAC isolates. The rising incidence of *C.auris* and *C.parapsilosis* as observed by us therefore holds greater significance.^[16] Our findings on species distribution of candida isolates were similar to the study on blood stream fungal isolates involving a large no. of countries in Asia-Pacific region,^[17] (2016) and an Equadorian study,^[18] (2020) and an Algerian study(2021) as well.^[19] *Candida albicans* isolates in this study remarkably showed highest percentage of resistance to

Amphotericine B which is not frequently observed worldwide till now.^[20] Mutations in the ERG3 gene that encodes a C-5 sterol desaturase, an enzyme involved in ergosterol biosynthesis, thereby lowering the concentration of ergosterol in the fungal membrane is mostly responsible for resistance to Amphotericine B.^[21] An increase in catalase activity, with decreased susceptibility to oxidative damage is also thought to render Amphotericine B ineffective.^[22] A few azole resistant strains showed cross-resistance to amphotericine B as well indicating the importance of selecting a new class of drug with a different target molecule as the therapeutic choice for drug resistant invasive candidiasis cases.^[23] Other than amphotericine B and fluconazole, flucytosine and Caspofungin also exhibited resistance though in less number of cases. Flucytosine monotherapy should therefore be avoided to prevent further development of resistance.^[24]

Caspofungin, a rapidly acting Echinocandin with very Low MIC for *C.albicans* and considered to be fungicidal in vitro is being increasingly prescribed for esophagitis caused by Fluconazole resistant *C.albicans*.^[25-27] Therefore, Caspofungin resistance is alarming as not only a few of the Fluconazole resistant isolates are showing complete or intermediate resistance to Caspofungin in-vitro but increasing MIC values are observed in few Fluconazole sensitive isolates as well.

Resistance of NAC against both Azoles and Echinocandin are increasing probably due to an increasing trend of using either of them as first line therapy in cases of invasive infections. *C.krusei* is intrinsically resistant to Fluconazole.^[28] *C.glabrata* isolates in our study showed higher proportion of resistance towards Fluconazole, Amphotericine B and Caspofungin though they remain highly susceptible against Voriconazole or Micafungin supporting a worldwide trend of increasing resistance of *C.glabrata* towards all first line antifungals. *C.auris* isolates were mostly resistant towards Fluconazole and one of the isolate was pan-resistant being in-vitro susceptible to Micafungin only.^[29,30] *Candida tropicalis*, the commonest NAC species isolated was in-vitro susceptible to most of the antifungals. The alarming finding though was significant proportion of *C.tropicalis* isolates being resistant to both amphotericine B and Fluconazole. The *Candida parapsilosis* isolates in our study showed a higher proportion of in-vitro resistance towards Amphotericine B.^[31]

Echinocandins namely Micafungin was the only drug to show 100% efficacy in this study against all *Candida* isolates. These drugs have now been used extensively as first line therapy in invasive and nosocomial candidiasis to reduce mortality. Judicious use of these drugs is therefore the need of the hour to prevent rapid emergence of resistance.

Limitation of the study: Vulvo-vaginal candidiasis cases could not be included in this study. Molecular study specially to determine the surge in

amphotericine B resistance in the *albicans* group could not be carried out.

CONCLUSION

Antifungal stewardship needs to be extensively planned and implemented specially taking account of the need for prophylactic and empirical therapy for the immunocompromised patients to reduce unnecessary use of antifungals as well as to reduce the cost of health care and prevent emergence of further drug resistance. Evaluation of different commercially available fungal marker detection tests in this regard can also be sought for.

REFERENCES

1. Riera FO, Caeiro JP, Angiolini SC, Vigezzi C, Rodriguez E, Icely PA, Sotomayor CE. Invasive Candidiasis: Update and Current Challenges in the Management of This Mycosis in South America. *Antibiotics* (Basel). 2022 Jun 30;11(7):877. doi: 10.3390/antibiotics11070877. PMID: 35884131; PMCID: PMC9312041.
2. Taei, M., Chadeganipour, M. & Mohammadi, R. An alarming rise of non-*albicans* *Candida* species and uncommon yeasts in the clinical samples; a combination of various molecular techniques for identification of etiologic agents. *BMC Res Notes* 12, 779 (2019). <https://doi.org/10.1186/s13104-019-4811-1>
3. Ortiz B, Aguilar K, Galindo C, Molina L, Fontecha G. *Candida* species isolated from clinical samples in a tertiary hospital in Honduras: Where is *Candida auris*? *Curr Med Mycol*. 2022 Sep;8(3):1-8. doi: 10.18502/cmm.8.3.11212. PMID: 37051554; PMCID: PMC10084484.
4. Megri Y, Arastehfar A, Boekhout T, Daneshnia F, Hörtnagl C, Sartori B, Hafez A, Pan W, Lass-Flörl C, Hamrioui B. *Candida tropicalis* is the most prevalent yeast species causing candidemia in Algeria: the urgent need for antifungal stewardship and infection control measures. *Antimicrob Resist Infect Control*. 2020 Apr 7;9(1):50. doi: 10.1186/s13756-020-00710-z. PMID: 32264966; PMCID: PMC7140370.
5. Reda NM, Hassan RM, Salem ST, Yousef RHA. Prevalence and species distribution of *Candida* bloodstream infection in children and adults in two teaching university hospitals in Egypt: first report of *Candida kefyr*. *Infection*. 2023 Apr;51(2):389-395. doi: 10.1007/s15010-022-01888-7. Epub 2022 Aug 26. PMID: 36018493; PMCID: PMC10042939.
6. Bhattacharjee P. Epidemiology and antifungal susceptibility of *Candida* species in a tertiary care hospital, Kolkata, India. *Curr Med Mycol*. 2016 Jun;2(2):20-27. doi: 10.18869/acadpub.cmm.2.2.5. PMID: 28681016; PMCID: PMC5490301.
7. Verma AK, Prasad KN, Singh M, Dixit AK, Ayyagari A. Candidaemia in patients of a tertiary health care hospital from north India. *Indian J Med Res*. 2003;117:122-8.
8. Bilal H, Zhang D, Shafiq M, Khan MN, Chen C, Khan S, Wang Q, Cai L, Islam R, Hu H, Zeng Y. 2023. Six-Year Retrospective Analysis of Epidemiology, Risk Factors, and Antifungal Susceptibilities of Candidiasis from a Tertiary Care Hospital in South China. *Microbiol Spectr* 11:e00708-23. <https://doi.org/10.1128/spectrum.00708-23>
9. Maheshwari, Monika, Kaur, Ravinder, Chadha, Sanjim, *Candida* Species Prevalence Profile in HIV Seropositive Patients from a Major Tertiary Care Hospital in New Delhi, India, *Journal of Pathogens*, 2016, 6204804, 8 pages, 2016. <https://doi.org/10.1155/2016/6204804>
10. Chouhan S, Kallianpur S, Prabhu KT, Tijare M, Kasetty S, Gupta S. Candidal Prevalence in Diabetics and its Species Identification. *Int J Appl Basic Med Res*. 2019 Jan-Mar;9(1):49-54. doi: 10.4103/ijabmr.IJABMR_259_18. PMID: 30820420; PMCID: PMC6385535.

11. Rodrigues CF, Rodrigues ME, Henriques M. *Candida* sp. Infections in Patients with Diabetes Mellitus. *J Clin Med*. 2019 Jan 10;8(1):76. doi: 10.3390/jcm8010076. PMID: 30634716; PMCID: PMC6352194.
12. Davenport JC. The oral distribution of candida in denture stomatitis. *Br Dent J*. 1970 Aug 18;129(4):151-6. doi: 10.1038/sj.bdj.4802540. PMID: 5272473.
13. Darwazeh A.M.G., Lamey P.-J., Samaranayake L.P., Macfarlane T.W., Fisher B.M., Macrury S.M., Maccuish A.C. The relationship between colonisation, secretor status and in-vitro adhesion of *Candida albicans* to buccal epithelial cells from diabetics. *J. Med. Microbiol*. 1990;33:43-49. doi: 10.1099/00222615-33-1-43
14. Rodrigues C.F., Rodrigues M., Henriques M. Susceptibility of *Candida glabrata* biofilms to echinocandins: Alterations in the matrix composition. *Biofouling*. 2018;34:569-578. doi: 10.1080/08927014.2018.1472244.
15. Guinea J. Global trends in distribution.....2014
16. Bhattacharjee P. Epidemiology and antifungal susceptibility of *Candida* species in a tertiary care hospital, Kolkata, India. *Curr Med Mycol*. 2016 Jun;2(2):20-27. doi: 10.18869/acadpub.cmm.2.2.5. PMID: 28681016; PMCID: PMC5490301.
17. Tan, Thean Yen & Hsu, Li & Alejandria, Marissa & Chaiwarith, Romanee & Chinniah, Terrence & Chayakulkeeree, Methee & Choudhury, Saugata & Chen, Yen-Hsu & Jong, Hee & Kiratisin, Pattarachai & Mendoza, Myrna & Prabhu, Dr & Supparatpinyo, Khuanchai & Tan, Ai & Phan, Xuan & Tran, Thi & Nguyen, Gia & Doan, Mai & Huynh, Van & Pham, Hung van. (2016). Antifungal susceptibility of invasive *Candida* bloodstream isolates from the Asia-Pacific region. *Medical Mycology*. 54. myv114. 10.1093/mmy/myv114.
18. Acosta-Mosquera Y, Tapia JC, Armas-González R, Cáceres-Valdiviezo MJ, Fernández-Cadena JC, Andrade-Molina D. Prevalence and Species Distribution of *Candida* Clinical Isolates in a Tertiary Care Hospital in Ecuador Tested from January 2019 to February 2020. *Journal of Fungi*. 2024; 10(5):304. <https://doi.org/10.3390/jof10050304>
19. Meradji A, Ranque S, Bachtarzi F, Mosbah N, Moulahem T. Incidence, Species Distribution, and Antifungal Susceptibility of *Candida* Bloodstream Infections in a Tertiary Algerian Hospital. *Biology and Life Sciences Forum*. 2024; 31(1):30. <https://doi.org/10.3390/ECM2023-16684>
20. Costa-de-Oliveira S, Rodrigues AG. *Candida albicans* Antifungal Resistance and Tolerance in Bloodstream Infections: The Triad Yeast-Host-Antifungal. *Microorganisms*. 2020; 8(2):154. <https://doi.org/10.3390/microorganisms8020154>
21. Kelly, S.L.; Lamb, D.C.; Kelly, D.E.; Manning, N.J.; Loeffler, J.; Hebart, H.; Schumacher, U.; Einsele, H. Resistance to fluconazole and cross-resistance to amphotericin B in *Candida albicans* from AIDS patients caused by defective sterol delta5,6-desaturation. *FEBS Lett*. 1997, 400, 80-82. [Google Scholar] [CrossRef] [Green Version]
22. Sokol-Anderson, M.L.; Brajtburg, J.; Medoff, G. Amphotericin B-induced oxidative damage and killing of *Candida albicans*. *J. Infect. Dis*. 1986, 154, 76-83. [Google Scholar] [CrossRef]
23. Helmerhorst EJ, Reijnders IM, van't Hof W, Simoons-Smit I, Veerman EC, Amerongen AV. Amphotericin B- and fluconazole-resistant *Candida* spp., *Aspergillus fumigatus*, and other newly emerging pathogenic fungi are susceptible to basic antifungal peptides. *Antimicrob Agents Chemother*. 1999 Mar;43(3):702-4. doi: 10.1128/AAC.43.3.702. PMID: 10049295; PMCID: PMC89188.
24. Jeniel E. Nett MD, PhD, David R. Andes MD. Fungal infections. *Infectious Disease Clinics of North America*, 2016
25. C. Kutler, B. Koll, B. Raucher, and B. Saltznab, 40th Annu. Meet. IDSA, abstr. 350, 2002).
26. Ernst, E. J., M. E. Klepser, and M. Pfaller. 2000. Postantifungal effects of echinocandin, azole, and polyene antifungal agents against *Candida albicans* and *Cryptococcus neoformans*. *Antimicrob. Agents Chemother*. 44:1108-1111.
27. Hernandez S, López-Ribot JL, Najvar LK, McCarthy DI, Bocanegra R, Graybill JR. Caspofungin resistance in *Candida albicans*: correlating clinical outcome with laboratory susceptibility testing of three isogenic isolates serially obtained from a patient with progressive *Candida* esophagitis. *Antimicrob Agents Chemother*. 2004 Apr;48(4):1382-3. doi: 10.1128/AAC.48.4.1382-1383.2004. PMID: 15047549; PMCID: PMC375251.
28. Pappas PG, Kauffman CA, Andes DR, Clancy CJ, Marr KA, Ostrosky-Zeichner L, Reboli AC, Schuster MG, Vazquez JA, Walsh TJ, Zaoutis TE, Sobel JD. Executive Summary: Clinical Practice Guideline for the Management of Candidiasis: 2016 Update by the Infectious Diseases Society of America. *Clin Infect Dis*. 2016 Feb 15;62(4):409-17. doi: 10.1093/cid/civ1194. PMID: 26810419.
29. Chesdachai S, Yetmar ZA, Ranganath N, Everson JJ, Wengenack NL, Abu Saleh OM. Antifungal Susceptibility Pattern of *Candida glabrata* from a Referral Center and Reference Laboratory: 2012-2022. *Journal of Fungi*. 2023; 9(8):821. <https://doi.org/10.3390/jof9080821>
30. Deshkar S, Patil N, Amberkar S, Lad A, Siddiqui F, Sharan S. Identification and Antifungal Drug Susceptibility Pattern of *Candida auris* in India. *J Glob Infect Dis*. 2022 Nov 1;14(4):131-135. doi: 10.4103/jgid.jgid_44_22. PMID: 36636301; PMCID: PMC9831210.
31. Branco J, Miranda IM, Rodrigues AG. *Candida* parapsilosis Virulence and Antifungal Resistance Mechanisms: A Comprehensive Review of Key Determinants. *J Fungi (Basel)*. 2023 Jan 5;9(1):80. doi: 10.3390/jof9010080. PMID: 36675901; PMCID: PMC9862255.